#### Edited wheat plants with CRISPR/Cas9













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# **INTRODUCTION**

Today wheat is the most cultivated cereal in the world. in our project we have used CRISPR/Cas (Clustered regularly interspaced short palindromic repeats ) which is a family of DNA sequences found in the genome of prokaryotic organisms and protein editing nucleases associated with these sequences (Cas) that cut DNA guided by RNA sequences and thus edit pieces of DNA.



- Gluten proteins are responsible of the breadmaking quality of wheat but also of some important pathologies.
- The main gluten proteins are: gliadins and glutenins.
- Gliadins are primarily responsible for celiac disease.

## **OBJETIVES**

The main objective of this work is to use biotechnological techniques, CRISPR/Cas vectors to edit wheat gliadin genes, to test the efficiency of the RNA guides for generating wheat lines with gliadin genes mutated.



## THEORETICAL FRAMEWORK

**Gluten:** The main storage group of grain proteins found in wheat. Being proteins they must be digested and broken down into small fractions. Some people have trouble digesting it causing damage.

**Celiac disease** is an autoimmune genetic disease in which there is a permanent intolerance to gluten.

In vitro culture: growing a plant in a Petri dish, containing nutrients which are provided, and specific environmental conditions are simulated.



#### THEORETICAL FRAMEWORK

**Cas9:** DNA endonuclease enzyme directed by a guide RNA, capable of identiying DNA and cutting both DNA strands, inactivating it. CRISPR/Cas is used in this research to prevent the synthesis of some proteins, which are harmful to people with celiac disease, taking advantage of the DNA repair pathways that are activated after DNA cutting





#### THEORETICAL FRAMEWORK

**Particle bombardment:** a technique used to introduce plasmids containing the DNA of some genes into plant cells. Gold particles, coated with plasmid DNA, are used to disrupt the cell wall so that foreign DNA enters the nucleus of the cell.

**PCR** – A technique that allows the rapid production of millions of replicates of a specific segment of DNA. What is needed is a DNA polymerase, nucleotides, primers, magnesium, and short fragments of synthetic DNA.





#### **MATERIALS AND METHODS**





Microcentrifuge

**Electrophoresis cuvettes** 

**Thermal cycler** 

Flowhood chamber



Micropipettes

Petri dishes

#### **DESIGN OF EXPERIMENTAL TOOLING**

- 1. Isolation of scutella.
- 2. Bombardment with CRISPR/Cas9.
- 3. In vitro culture:
  - Embryogenesis
  - Plant regeneration
  - Plan in soil
- 4. PCR of Cas9 and Electroforesis.
- 5. Analyze results.



### 1st SESSION

The first session (23/11/22) we were explained what the work would consist of and proceeded to prepare the mixture of DNA and gold to carry out the bombardment of the previously isolated scutellum. With this, we introduced the Cas9 gene and the guide RNAs into the wheat scutellum cells.

## **2nd SESSION**

The second session (11/01/23) the embryogenic capacity of the wheat scutellum was determined and they were transferred to another medium to induce regeneration. At all times, work was carried out in a laminar flowhood chamber





### **3rd SESSION**

In the third session (08/02/23) the scutellum were also transferred to another new plate, or to a magenta or to pots, depending on the level of development of the explants/plantlets. In addition, DNA was extracted for PCR to detect the presence of the Cas9 gene.





### 4th SESSION

In the fourth session (08/03/23) more plants were transferred to magenta or soil pots. Afterwards, in the IAS meeting room, the data were analysed, the tasks for the report were distributed and the design of the poster was worked on.

### **RESULTS**

**Table 1**. Efficiency of the regeneration and transformation of wheat scutella

Bombarded	Regenerated	Cas9 + <u>plants</u>	Regeneration	Transformation
scutella	plants		efficiency	efficiency
262	49	10	18.7	3.8





30%









**Figure 2.** Identification of regenerated plants. Results of amplification of the gene encoding the Cas9 protein in regenerated plants.

Alpha 19Alpha 5Alpha 20Alpha 7Alpha 20Alpha 6H2OAlpha 8H2OAlpha 10Alpha 11Alpha 11+Alpha 12+Alpha 13+Alpha 14+Alpha 15+Alpha 16+Alpha 16+Alpha 16+Alpha 16+Alpha 16+Alpha 16+Alpha 16+Alpha 16+Alpha 16+Alpha 16Alpha 17Alpha 18

**6** 4

Alpha Alpha Alpha

ω

α

Alpha

**Figure 3.** A-PAGE gel of wheat gliadins from lines transformed with the pSSLAlpha9 vector.

pSSLApha9

WT ---

## CONCLUSIONS

- 1. Over 60% of wheat scutella produce somatic embryos.
- 2. The in vitro selection system is highly efficient and allows to regenerate 49 plants (18.7%)
- 3. Around 20% of the plants analyzed contained the gene that codes for Cas 9.
- 4. The CRISPR/Cas9 system allows the editing of genes that code for the proteins responsible for triggering gluten intolerances.



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## END

## THANK YOU ALL FOR YOUR ATTENTION